

REVIEW

Interactions between Reactive Oxygen Species and Cancer: the Roles of Natural Dietary Antioxidants and their Molecular Mechanisms of Action

Ademola A Oyagbemi*, OI Azeez, AB Saba

Abstract

Reactive oxygen species (ROS) are natural products inevitably generated along with cellular metabolism. Due to their extreme reactivity, they can damage DNA, proteins and lipids. Dietary antioxidants have been shown to take part in cellular reduction-oxidation (redox) reactions in which they can act as either antioxidants (electron donors) or pro-oxidants (electron acceptors) depending on the physiological environment and general oxidative state. Organisms have developed efficient machinery and mechanisms to keep the production of ROS under tight control, these same mechanisms have also been found to regulate other intracellular processes. p53 is a sequence-specific transcription factor and critical tumour suppressor gene that is most frequently mutated in human cancer. Cancer, one of the leading causes of death worldwide, can now be ameliorated, blocked or reversed with ubiquitous polyphenolic and organosulphur compounds present in natural dietary antioxidants.

Key words: Reactive oxygen species - DNA damage - cancer - antioxidant phytochemicals

Asian Pacific J Cancer Prev, 10, 535-544

Introduction

Reactive oxygen species (ROS) are natural products inevitably generated along cellular metabolism (Lien et al., 2008). Due to their extreme reactivity, they can damage DNA, proteins and lipids. High levels of ROS have been shown to induce apoptosis as chronic high levels can promote vascular diseases such as arteriosclerosis (Lien et al., 2008). ROS include superoxide anion (O_2^-), hydrogen peroxide (H_2O_2) and highly reactive by-product of H_2O_2 with hydroxyl radicals (OH) that are capable of reacting with and damaging DNA, proteins and lipids (Rajeshwar, 1996).

Under normal conditions, cells utilize antioxidant defense systems to balance these toxic products (ROS) to keep the cells in a state of redox homeostasis (Lien et al., 2008). The deleterious effects of oxygen are said to result from its metabolic reduction to these highly reactive and toxic species (Buechter, 1988). Oxidative damage to cellular DNA has been reported to lead to mutation (Gulam & Haseeb, 2006) and this plays an important role in the initiation and progression of multistage carcinogenesis. Alterations in DNA such as base modification, rearrangement and miscoding of DNA base sequence, miscoding of DNA base sequence, gene duplication and subsequent activation of oncogenes are all known to be involved in initiating various cancers. Reactive oxygen species (ROS) and the related oxidative damage have also been implicated in the pathogenesis of various chronic

human diseases (Pincemail, 1997; Ames, 1993; Witztum, 1994; Halliwell, 1993).

ROS can cause tissue damage by reacting with lipids in cellular membranes, nucleotides in DNA (Ahsan et al., 2003) and sulphhydryl groups in proteins (Knight, 1995). Halliwell (1997) reported that free forms of ions are generated in the decomposition of iron-containing natural sources such as hemoglobin and ferritin. ROS can also be formed through lipid oxidation and photo-sensitizers when exposed to light (Boff and Min, 2002). Pro-oxidative enzymes such as lipo-oxygenase can generate free radicals (Spiteller, 2001) and significant amount of ROS can be generated through cross-linking/fragmentation of ribonucleoproteins (Waris & Alam, 1998). The relatively un-reactive superoxide anion radicals is converted by superoxide dismutase (SOD) into H_2O_2 which in turn take part in the "Fenton reaction" with transition metal ion (copper or iron) as catalysts to produce the very reactive hydroxyl radical (Aruoma, 1989; Halliwell & Gutteridge, 1990; 1992). Oxygen derived from free radicals have also been implicated directly or indirectly in a wide arrays of clinical disorders such as atherosclerosis, reperfusion injury, pulmonary toxicity, macular degeneration, cataracto-genesis, and cancer (Knight, 1995).

Dietary antioxidant has been shown to take part in cellular reduction-oxidation (redox) reactions in which they can act as either antioxidants (electron donors) or prooxidants (electron acceptors) depending on the physiological environment and general oxidative state

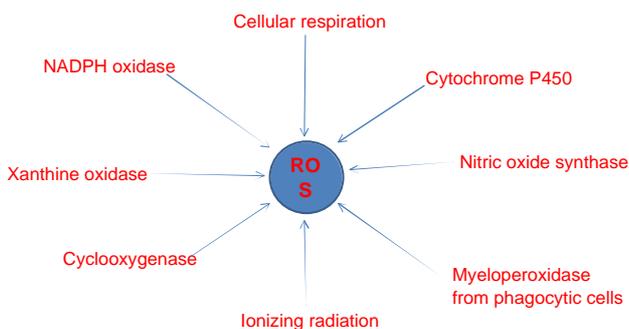


Figure 1. Sources of Reactive oxygen Species

(Schwart, 1992; Palozza, 2002). Flavonoids which act as antioxidants include epigallocatechin gallate (tea), quercetin e.g. from onion, red wine, and berries, genistein (soybean), and taxifolin from citrus fruits (Heim, 2002). Other classes of dietary phytochemicals have been shown to considerably attenuate ROS generation that can take part in redox reactions in addition to flavonoids include broad categories of carotenoids (IARC, 1998) and organosulfur compounds (Bianchini and Vainio, 2001).

Generation of ROS

ROS are formed via several mechanisms (see Figure 1) including (1) ionizing radiation on biological molecules, (2) as an unavoidable by-product during cellular respiration, and (3) synthesized by enzymes (NADPH oxidase and myeloperoxidase, from phagocytic cells to battle against bacterial infection (Martindale and Holbrook, 2002; Klaunig & Kamendulius, 2004; Poli et al., 2004). Fibronectin, a major component of extracellular matrix, has been reported to stimulate ROS production through activation of NADPH oxidase dependent and NADPH oxidase independent pathways and 5’lipoxygenase mediates these pathways (Mouad et al. 2005).

Under the conditions of normal metabolism the most important source of superoxide anion (O_2^-) is the mitochondrial electron transport chain, which leaks a few electrons directly onto O_2 as part of normal metabolism. It is estimated that 1 % to 3% of O_2 reduced in mitochondria is in the form of O_2^- Turrens (2003). This product comes from two sites, complex 1 (NADH dehydrogenase) and also complex III (ubiquinone-cytochrome c reductase), with the latter being the major source under normal conditions (Salvemini and Cuzzocrea, 2002).

Several enzymes also contribute to O_2^- production. One of the best characterized is xanthine oxidase (XO), which is present in the cytosol of many tissues and circulating blood or bound to glycosaminoglycan sites in the arterial wall (White, 1996). Normally, the enzyme acts as a dehydrogenase and transfers electrons to NAD^+ rather than O_2 , in ischemia reperfusion (Ullrich and Bachschmid, 2000; Mueller, 2005) or in sepsis (Mueller, 2005; Brandes et al., 1999) in which the active site of the enzyme is oxidized and the enzyme acts as an oxidase and produces O_2^- . The same is the case with a number of metabolically active enzymes as part of their normal function or when there is inadequate substrate. For example, cytochrome P450 enzymes in its reaction cycle can produce O_2^- as a

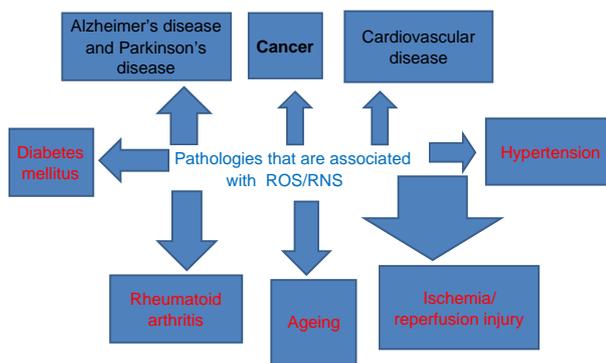


Figure 2. Diseases Associated with ROS and RNS

side reaction when they breakdown target molecules (Halliwell and Gutteridge, 1999).

Nitric oxide synthases, the family of enzymes that produce NO^- , produce O_2^- when the substrates L-arginine or co-factor tetrahydropteridines are insufficient (Mueller, 2005; Xia, 1996; and Xia, 1997)]. O_2^- can also be produced by cyclooxygenase as part of arachidonic acid metabolism. O_2^- even can be produced through auto-oxidation of molecules such as glyceraldehyde, $FMNH_2$, $FADH_2$, adrenalin, noradrenalin, dopamine and thiol containing molecules such as cysteine in the presence of O_2 (Salvemini and Cuzzocrea, 2002; Halliwell and Gutteridge, 1999). A number of diseases are associated with ROS or reactive nitrogen species (see Figure 2).

DNA damage and cancer

DNA damage by ROS has been accepted as a major cause of cancer according to Ames (1993). Increase coupled with deficient repair systems has been found to contribute significantly as major risk factors in diseases such as chronic hepatitis, cystic fibrosis, and various autoimmune diseases associated with increased risk of cancer (Hagen et al., 1994; Shimoda et al., 1994; Brown, 1995; Waris, 2005). ROS have been reported to be involved in modification of guanine, causing G-T transversions (Lunec, 2002). When critical genes such as oncogenes or tumour suppressor genes are affected, initiation/progression can result (Ames 1994; Moller, 1994). In human tumour, DNA base modifications including G to T transversions have been shown to be the most frequent mutations in the P53 suppressor gene (Harris and Hollstein, 1993). Elevated levels of modified bases in cancerous tissue have been reported to result from production of large amount of H_2O_2 which is characteristic of human tumour cells (Szatrowski & Nathan, 1991; Olinski, 1998). Activated carcinogens are known to exert their biological effects by forming covalent adducts with the individual nucleic acids of DNA or RNA. Similarly, reactive oxygen species (ROS), such as superoxide anions, hydrogen peroxide, and hydroxyl radicals, have been found to attack both DNA bases and the deoxyribosyl backbone of DNA (Bianchini et al., 2001).

DNA adducts particularly distort the shape of the DNA molecule, potentially causing mistranslation of the DNA sequence. Secondly, when the DNA replicates, an adducted base that persists unrepaired can be misread, producing mutations in critical genes, such as oncogenes and tumour suppressor genes. Thirdly, repair of bulky adducts can

result in breakages of the DNA strand, which can, in turn, result in mutations or deletions of genetic material (Knight, 1995). Numerous DNA repair pathways are known to exist and function to prevent the persistence of damage in DNA and are integral to the maintenance of genome stability and prevention of cancer (Martindale & Holbrook, 2002). Various DNA repair mechanisms include direct repair, base excision repair, nucleotide excision repair, double-strand break repair, and repair of inter-strand cross-links (Klaunig & Kamendulius, 2004).

DNA damage and p53

p53 is a sequence-specific transcription factor and critical tumour suppressor gene most frequently mutated in human cancer (Levine, 1997). p53 trans-activates genes that mediate apoptosis and has roles in DNA repair, senescence, and cell cycle arrest. There is enough compelling evidence that the primary physiologic role of p53 in DNA damage-induced apoptosis is to function as a transcriptional activator of genes encoding apoptosis effectors. p53 directly activates transcription of several genes encoding members of the Bcl-2 family, but it also mediates cell death through a variety of other mechanisms, including down-regulation of anti-apoptotic genes such as Map4 and survivin and up-regulation of pro-apoptotic genes such as Bax, IGF-BP3, DR5, Fas, and Apaf-1, as well as various other apoptosome components representing potential key therapeutic targets (Woods & Vousden, 2001; Hajra and Liu, 2004; Slee, 2004). p53 has also been shown to exhibit a direct apoptogenic role in the mitochondria, where it translocates and interacts with Bcl-xL and Bcl-2 proteins to induce mitochondrial permeabilization (Mihara, 2003). Moreover, p53 deficiency leads to inappropriate survival of cells with DNA damage and therefore predisposes individual to develop neoplasia.

Recently, p63 and p73 proteins have also been identified that bind p53 response elements and trans-activate p53-associated genes and, as a result, induce apoptosis. Furthermore, there is extrinsic overlap of p53 and multiple transcriptional targets, in which p53 can activate at least two proteins in the intrinsic pathway, including Bax and p53-apoptosis inducing factor (Harms, 2004). Reactive oxygen species have been strongly correlated with p53-mediated apoptosis. Upon over-expression of p53, ROS levels rise, and mitochondrial apoptosis is induced as aforementioned. Inhibition of ROS-mediated apoptosis has also been reported in smooth muscle cells (Johnson, 1996).

ROS and Cell signalling

ROS are potentially highly toxic molecules released during redox reactions and they are part of the basic chemical processes of life (Finkel, 1998; Finkel & Holbrook, 2000) (see Figure 3). Organisms have developed efficient machinery and mechanisms to keep the production of ROS under tight control, these also regulating other intracellular processes (Forman et al. 2002; Droge, 2002; Thannickal & Fanburg, 2000; Schafer & Buettner, 2001; Janssen-Heininger, 2000) or can activate

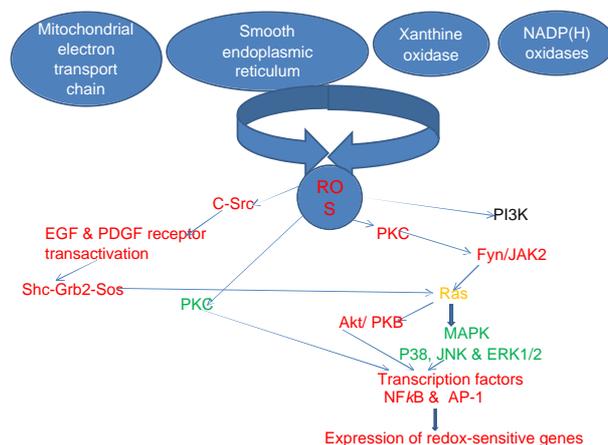


Figure 3. ROS-Mediated Mechanisms

diverse signaling pathways and affect arrays of various gene expressions. Lien et al., (2008) recently reported activation of Mitogen-activated protein kinases (MAPKs), phospholipase C-γ (PLC-γ) signaling, Protein Kinase C (PKC), p53 signaling, ataxia-telangiectasia-mutated (ATM) kinase, nuclear factor kappa B (NF-κB) and Jak/Stat pathway by ROS. The MAPK signaling pathways modulate gene expression, mitosis, motility, metabolism, and programmed cell death (Gulam and Haseeb, 2006).

A lot of evidence indicating that ROS and the redox state have a signaling role in bacteria and plants has been reported, but there was scanty evidence in mammalian cells until recently. For example, in bacteria, the transcription factor OxyR is redox sensitive. There is now an increasing number of examples of ROS-based signaling in animals, including protein tyrosine phosphatase 1B (PTP-1B) (Lee, 2002), thioredoxin (Saitoh et al., 1998), SERCA2 (Adachi et al., 2004) and Ras (Adachi et al., 2004). Nitric oxide (NO·) is a well-characterized radical that has a major role in normal physiological functions. This radical has a central role in the regulation of vascular tone, nerve function and immune regulation. Even the potentially toxic by-products of NO· and O₂⁻, OONO·, has been shown to contribute to control of vascular tone (Adachi et al., 2004).

Cohen and coworkers found that NO· induced dilatation occurs by the production of low concentrations of OONO·, which directly stimulates the sarco/endoplasmic reticulum calcium (Ca²⁺) ATPase (SERCA) to decrease intracellular Ca²⁺ and thereby produce vasodilatation. This occurs by reversible S-glutathiolation of the thiol of a cysteine molecule on SERCA. Thus, by removing O₂⁻ and preventing the formation of OONO·, superoxide scavengers actually blocked NO-induced vascular relaxation. However, high levels of oxidative stress, including high concentrations of OONO·, have been reported to result in irreversible oxidation of key thiols and prevented normal NO· induced relaxation. Endothelial and neuronal cells that use NO for signalling produce NO in small amounts, whereas macrophages and neutrophils that use NO to attack invading organisms produce it in large amounts. In the same vein, the NAD(P)H oxidase in phagocytic cells produces large quantities of O₂⁻, whereas the NAD(P)H oxidases in non-phagocytic cells produce much smaller amounts, consistent with a signalling role.

The MAPK kinase signaling cascades include

extracellular signal-related protein kinases (ERKs), JNKs/stress-activated protein kinases (SAPKs), and p38 kinases. The ERKs transmit signals initiated by growth promoters, including EGF, PDGF, and fibroblast growth factor (FGF) and may ultimately foster cell growth and survival (Bode and Dong, 2004). The polyphenols; curcumin, EGCG, and resveratrol downregulate phosphorylation and ligand binding of growth factor receptors including EGF, FGF, and PDGF (Manson, 2005). Consequently, this quenches MAPK signaling, transcription factor activation (i.e., AP-1), and ultimately gene expression. It is noteworthy that many cells require such signals to avoid apoptosis, and, as a result, interruption of this signaling encourages induction of apoptosis in many cell types. For example, the indirect inhibition of PI3-Akt anti-apoptotic signals might contribute to cell death through modulation by diet (Chen and King, 2005). The MAPKs are activated by translocation to the nucleus, where they phosphorylate numerous substrates, including the transcription factors AP-1 and NF- κ B. Activation of both are linked to carcinogenesis and promotion (Bode and Dong, 2004).

Indeed, numerous mutations can occur in tumour suppressor genes involved in induction of apoptosis, and these include p53, p19, ARF, Rb, PTEN, TRAIL, and CD95/Fas (Johnstone, 2002). Numerous oncogenes may also be activated through mutation to inhibit or circumvent the inherent controls of apoptosis, and critical oncogenes involved include Bcl-2, MDM2, IAPs, NF- κ B, Akt, PI3K, Ras, Myc, and FLIP (Johnstone, 2002). Blocking the expression of these genes, and in particular oncogenic ras, is currently an active pharmacological approach for cancer therapy (Adjei, 2001). Mutations in genes that regulate apoptotic pathways are common in most cancers, emphasizing their importance for DNA damage (Sun et al., 2004).

Many dietary agents can affect cellular signaling. Resveratrol stimulates complex formation between p53, ERK, and p38 kinase, with enhanced phosphorylation, stabilization, and activation of p53 in epidermal cells (Roemer and Mahyar-Roemer, 2002). Indole-3-carbinol potently inhibits signaling through protein kinase B and binding of NF- κ B to DNA (Howells et al., 2002). Indole-3-carbinol and DIM also inhibit the MAPK pathway, which may inhibit cancer cell survival. Curcumin reduces the activity of p38 MAPK, and EGCG inhibits tyrosine kinase and MAPK activation in transformed cells but not in normal cells. Capsaicin, a principal pungent ingredient of hot red and chili peppers, markedly activates JNK-1 and p38 MAPK signaling in Ha-ras-transformed human breast epithelial cells (Hail, 2003). In cells with mutated oncogenic Ha-ras, green and black tea polyphenols potently inhibited ERK phosphorylation and AP-1 activity (Chung et al. 1999). Allyl ITC (AITC), BITC, and PEITC increased activity of JNK in HL-60 cells. MAPK, ERK, and p38 kinase were activated by PEITC in HT29 and PC3 cells. Similarly, BITC has been shown to activate p38 kinase in human head and neck squamous cell carcinoma lines (Zhang, 2004). In the same vein, garlic compounds, DADS potentially induced ROS and JNK, S-allylmercaptocysteine induced JNK-1 activation and jun kinase activity, as well as ajoene activated MAPKs (JNK,

p38, ERK1/2) in different cell types (Wu et al., 2005).

The role of ROS in the signaling of a number of growth factors has also been well established. A superb example is the role of ROS in angiotensin signaling as established by Griendling and co-workers (Griendling et al., 2000; Griendling et al. 1994; Ushio-Fukai et al., 1999 and Zafari et al., 1998). This group of researchers showed that exposure of vascular smooth muscle to angiotensin II results in smooth muscle growth that is dependent upon increased production of O₂⁻ by NAD(P)H oxidase and its subsequent dismutation to H₂O₂. H₂O₂ then activates downstream prosurvival pathways with resultant vascular hypertrophy. Other growth factors such as platelet derived growth factor (PDGF) have been shown to have similar signaling mechanisms (Sundaresan et al., 1995) ROS also play a significant role in the intracellular signaling of tumour necrosis factor- α (TNF- α) (Matsubara & Ziff, 1986; Ferro et al., 1997; Ferro et al., 19998; De Keulenaer et al., 19998; Murphy et al., 1992; Phelps et al., 1995; Goode & Webster, 1993; Lum & Roebuck, 2001 & Demling et al., 1986) and this probably occurs through O₂⁻ produced by NAD(P)H oxidase and regulation of the transcriptional activity of NF κ B. Also, it has recently been shown that lipopolysaccharide activation of Toll-like receptor 4 increases O₂⁻ production by NAD(P)H oxidase and this too leads to NF κ B activation (Park et al., 2004).

Properties of some Chemopreventive Agents and their Molecular Mechanisms of Action

Resveratrol

There is mounting evidence in the literature that Resveratrol is a promising natural compound for prevention and treatment of a variety of human cancer (Fulda and Debatin, 2006). Resveratrol, trans-3, 5, 41-trihydroxy-trans-stilbene, is a phytoalexin produced by plants, and the skin of red grapes. Resveratrol affects all three stages of carcinogenesis, namely initiation, promotion and progression (Mohammad, 2007). Resveratrol has been shown to block carcinogen activation and subsequent DNA damage by suppressing induction of Phase I metabolizing enzymes (Kudu and Surh, 2005).

The anticarcinogenic effect of resveratrol has been found to be closely associated with its antioxidant activity, through which it inhibits cyclooxygenase, hydroperoxidase, protein kinase C, Bcl-2 phosphorylation, Akt/protein kinase B, focal adhesion kinase, NF- κ B, matrix metalloproteinase-9, and cell cycle regulators. Exposure of normal cells to resveratrol results in the activation of a series of upstream kinases such as ERK, JNK and P13K, with dissociation of nuclear factor related erythroid factor 2 (NrF2) from its inhibitory counterpart keap1. Free NrF2 translocates to the nucleus, where it regulates transcriptional activation of genes encoding phase II detoxification/antioxidant enzymes (Kudu and Surh, 2005). Resveratrol has been reported to modulate diverse signal transduction pathways resulting in the blockade of carcinogen activation and enhancement of detoxification, inhibition of inflammation, cell proliferation, and apoptosis (Kudu and Surh, 2005). Antioxidant activity of resveratrol has been attributed to

its ability to inhibit H_2O_2 production, myeloperoxidase activity and restoration of glutathione levels and activity of superoxide dismutase (Jang & Pezzuto, 1998). Resveratrol has also been reported to inhibit COX-2 at both transcriptional and post-transcriptional levels (Subbaramaiah et al., 1998). It also down regulates the expression of both COX-1 and COX-2 mRNA transcripts in NMBA-induced esophageal tumour in F34A rats (Li et al., 2002) and inhibition of LPS, TPA or H202-induced mobilization of arachidonic acid and expression of COX-2 which results in decrease PGE2 production (Martinez and Moreno, 2000). Resveratrol has been shown to inhibit the expression of various Cyclins (e.g. D1, D2 and E) as well as the expression and catalytic activities of Cyclin dependent kinases (CDK) -2, -4 and -6 suggesting that resveratrol-induced up regulation of p21WAF-1/CIP-1 may inhibit the formation of Cyclin-cdk complexes thereby imposing artificial check points at G1/S transition of the cell cycle (Ahmad et al., 2001) and induces stabilization and activation of p53 (Roemer and Mahyar-Roemer, 2002).

Lycopene

Lycopene is known to be an effective scavenger of reactive oxygen species, including singlet oxygen (O_2^1) and other excited species (Stahl et al., 1998 & Woodall et al., 1997). Lycopene is the strongest singlet oxygen quencher as well as potent antioxidant compared to other carotenoids. Lycopene is mainly contained in tomatoes and high concentration is found in processed tomato products like ketchup, tomato sauce/juice as well as in red coloured fruits like water melon and guava. Intake of lycopene has been inversely linked to the incidence of prostate cancer (Giovannucci et al., 2002; Sesso et al., 2004). Lycopene is the main carotenoid in tomatoes and tomato-products and is responsible for the red colouration of tomatoes (Hwang and Lee, 2006). Suggested inhibitory mechanisms for lycopene include enhancing gap junction intercellular communication (Krutvoskikh et al., 1997), cell cycle arrest (Hwang and Bowen, 2000), suppression of tumour cell proliferation and apoptosis (Kim et al., 2001). Lycopene is one of the most potent antioxidants (Miller et al., 1996) and has been suggested to prevent carcinogenesis and atheriogenesis by protecting critical macromolecules including lipids, low-density lipoproteins (LDL), proteins and DNA (Rao & Agarwal, 1998; Pool-Zobel, 1997). Lycopene, because of its high number of conjugated double bonds, exhibits higher singlet oxygen quenching ability compared to β -carotene or α -tocopherol (Dimasco et al., 1989). Lycopene in small doses reduced the N-methylnitrosourea (MNU) induced development of aberrant crypt foci (ACF) in the colon of Sprague-Dawley rats (Narisawa et al., 1998), dimethylbenzanthracene (DMBA)-induced mammary tumour (Sharoni et al., 1997) and diethylnitrosamine (DEN) induced liver pre-neoplastic foci in rats. Ingestion of tomato juice has been shown to inhibit the development of N-butyl-N-(4-hydroxybutyl) nitrosamine (BBN)-induced development of urinary bladder in transitional cell carcinomas in male Fischer 344 rats (Okajima et al., 1998), protection against azoxymethane (AOM)-induced colonic pre-neoplastic lesions (Jain et al., 1999). Similarly, lycopene has also

been shown to protect against microcystin-induced mouse hepatocarcinoma by suppressing the phosphorylation of regulatory proteins such as retinoblastoma gene protein product and arresting cells in the Go/G1 phase of cell cycle similar to P53 function (Matsushima et al., 1995)

Curcumin

Curcumin, a widely used spice and colouring agent in food, has been shown to possess potent antioxidant, antitumour promoting and anti inflammatory properties *in vivo* and *in vitro* (Motterlini et al., 2000). Copper complex of Curcumin was found to show promising superoxide dismutase (SOD) activity with comparable free radical scavenging ability and an improved antioxidant efficacy; meaning that Curcumin could also act as antioxidant enzyme. Curcumin was recently reported to protect against dimethyl nitrosamine (DMN)-induced hepatotoxicity through antioxidant response element (ARE)-driven induction of heme-oxygenase-1 [HO-1] expression (Farombi et al., 2008). The increased haeme-oxygenase activity is an important component of Curcumin-mediated cyto-protection against oxidative stress. Pretreatment of human colonic epithelial cells with Curcumin has been shown to inhibit TNF- α -induced cyclooxygenase -2 (COX-2) gene transcriptions and NF- κ B activation and inhibition of IKB degradation by down regulation of NF- κ B-inducing kinase (NIK) and IKB kinase (IKK). Curcumin was shown to inhibit IKB phosphorylation in human multiple myeloma cells (Bharti et al., 2003) and murine melanoma cells through suppression of IKK activity, and this taken together contributes significantly to its antiproliferative, pro-apoptotic and/or anti-metastatic activities.

Capsaicin

Hot red chili pepper which belongs to the plant genus capsicum is widely consumed all over the world. Nigeria is not an exception. 8,-methyl-N-Vanillyl-6-nonenamide is the principal phenolic compound in capsaicin. Chemopreventive properties of capsaicin have been reported (Surh and Lee, 1995; Surh, 2002; Philip & Kundu, 2003). Topical application of capsaicin has been shown to inhibit PMA-induced mouse-skin tumour formation (Park et al., 1998) and activation of NF- κ B (Han et al., 2002). The anti-proliferative property of capsaicin has been ascribed to its ability to induce apoptosis (Lee et al., 2000; Jung et al., 2001). Generation of reactive oxygen species (ROS) is one of the mechanisms by which capsaicin induce apoptosis in tumour cells. Macho et al (1998) reported induction of apoptosis in cultured Jurkat cells through generation of reactive oxygen species (ROS) and rapid activation of C-JUN NH₂-terminal kinase (JNK).

Capsaicin also causes G1 arrest of endothelial cell through down-regulation of cyclin D1 and vascular endothelia growth factor (VEGF) induced angiogenic signaling pathways. Cyclin D1 is required for the activity of cyclin-dependent kinase 4(CDK4) which phosphorylates RB (tumour suppressor gene), thereby releasing E2F to mediate the transition of G1 to S, which in turn leads to DNA synthesis and cell cycle progression (Jeong, 2004) and this pathway is blocked by capsaicin. It

has been documented that other anti-angiogenic molecules such as endostatin and Curcumin also suppress (Retinoblastoma gene) Rb phosphorylation and DNA synthesis of endothelial cells through down-regulation of cyclin D1 (Mukhopadhyay et al., 2002; Hanai et al., 2002). In fact, capsaicin was reported to block the downstream event of Vascular endothelial growth factor (VEGF) induced KDR/Flk-1 signaling such as activation of p38 mitogen activated protein kinase and p125 FAK tyrosine phosphorylation that are required for the mitogenic activity of VEGF in endothelial cells (Bernatchez et al., 1999; Zachary & Glick, 2001 & Davis-Smyth, 1996). Capsaicin modulates the activities of pro-inflammatory mediators and the intracellular signaling cascades. Researches are currently being carried out to unravel the molecular mechanisms of action and other signaling pathways that are associated with capsaicin anti tumour effects.

Sulphoraphane

Sulphoraphane (SF) is a naturally occurring isothiocyanate which has a potent anticarcinogenic capability in experimental animal models. This isothiocyanate is quantitatively resident in broccoli and has been shown to be a potent inducer of phase 2 detoxification enzymes e.g. glutathione S-transferases (GSTs), and also known to block metabolic activation of chemical carcinogens in experimental animal models (Zhang et al., 1992). Other mechanisms associated with antiproliferative and chemopreventive activity of sulphoraphane include induction of apoptosis, cell cycle arrest, anti-inflammation and ability to inhibit phase I enzymes that might be involved in activation of chemical carcinogens to ultimate carcinogens (Chung et al., 2000; Zhang, 2000). It was recently shown that SF, as well as some of its analogues, rapidly accumulated in all cell lines tested, and its intracellular concentrations can reach millimolar levels (Zhang and Talalay, 1998; Zhang, 2000). SF appeared to enter cells freely, but was almost entirely conjugated with GSH in cells (Zhang, 2000; 2001). Results from investigations revealed that cellular GSH was the principal driving force for accumulation while cellular GST further enhances such accumulation (Zhang and Talalay, 1998; Zhang, 2001). This mechanism is consistent with the fact that SF undergoes spontaneous conjugation with GSH under mild conditions to give rise to the corresponding dithiocarbamate (GS-SF) and that the reaction is accelerated by GST (Kolm et al., 1995). Preliminary evidence suggests that the accumulated GS-SF may be further metabolized, such as by binding to cellular proteins and the formation of other dithiocarbamate metabolites (Zhang, 2000; Kassahun et al., 1997). Cellular accumulation of SF appears critical for its anticarcinogenic activity, related to its induction of many Phase 2 detoxication enzymes in cultured cells (Zhang & Talalay, 1998; Ye & Zhang, 2002).

Caffeic acid phenethyl ester (CAPE)

Caffeic acid phenethyl ester (CAPE) is an active component of propolis from honeybee hives (honeybee resin). CAPE has been shown to have anti-inflammatory, anti-carcinogenic and immunomodulatory properties

(Grunberger et al., 1988 ; Abdel-Latif et al., 2005). CAPE was shown to suppress acute inflammation (Orban et al., 2000), acts as specific inhibitor of NF- κ B (Natarajan et al., 1996) and inhibition of nuclear factor of activated cells (NFAT) and activator protein-1 (AP-1) nuclear binding and activation. CAPE also alters the redox state, induces apoptosis, suppresses lipid peroxidation and acts as an antioxidant (Kimura et al., 1984; Chiao et al., 1995; Laranjinha et al., 1995) and inhibits COX-2 expression (Kim et al., 2001). The inhibitory potentials of CAPE on different transcription factors provide direction and deep insight into the nuclear mechanism(s) of actions underlying the chemopreventive properties of CAPE. It has anti-inflammatory, anti-viral, anti-mitogenic, anti-carcinogenic, and immunomodulatory effects (Ozyurt et al., 2004; Song et al., 2000). It has also been reported that CAPE exhibits antioxidant activity and inhibits lipoxygenase activities, protein tyrosine kinase, and NF κ B activation (Natarajan et al., 1996; Song et al., 2002). The possibility that CAPE displays pharmacological activity by inhibiting the release of arachidonic acid and the enzyme activities of cyclooxygenase (COX)-I and COX-II have also been extensively discussed (Song et al., 2002).

Diallyl Sulphide (DAS)

Medicinal use of garlic has been dated back to antiquity. Diallyl sulphide is the fat soluble bioactive compound in garlic. DAS is an antioxidant and anti-inflammatory agent (Kalayarasan et al., 2008). DAS has been shown to reduce bleomycin-induced activation of inducible nitric oxide synthase (iNOS) and NF κ B and decrease the augmented levels of the early inflammatory cytokines, tumour necrosis factor alpha (TNF- α) and interleukin I beta (IL-1 β) in lung tissues (Kalayarasan et al., 2008). Thejass and Kuttan (2007) recently suggested that antiangiogenic activity of DAS can be related to its negative regulation of pro-angiogenic factors such as VEGF and proinflammatory cytokines and positive regulation of antiangiogenic factors such as IL-2 and TIMP. Apoptotic activity of DAS can be adduced from the work of Sriram (2008) which include increased production of ROS, cell cycle arrest, decreased cell proliferation and induction of apoptosis. DAS also promotes the expression of caspase-3 and suppresses the activity of extracellular regulatory kinase-2 (ERK-2). Other antioxidant and anticancer activities of DAS have also been reported by various authors (Green et al., 2007a; Green et al., 2007b; Arora et al., 2006; Thomas et al., 2004).

Conclusions

The use of chemopreventive agents in natural products has become the mainstay and novel sources of alternative therapy in our contemporary days. Their chemopreventive potentials which include antioxidant, antiproliferative, anticancer, antiangiogenic properties are now being utilized for human benefit to alleviate various forms of degenerative diseases including cancer. Cancer, one of the leading causes of death worldwide can now be ameliorated, blocked or reversed with these polyphenolic and organosulphur compounds present in these phytochemicals.

References

- Abdel-Latif MM, Windle HJ, Homasany BS, Sabra K, Kelleher D (2005). Caffeic acid phenethyl ester modulates Helicobacter pylori-induced factor-kappa B and activator protein-1 expression in gastric epithelial cells. *Br J Pharmacol*, **146**, 1139-47.
- Adachi (2004). S-Glutathiolation by peroxynitrite activates SERCA during arterial relaxation by nitric oxide. *Nat Med*, **10**, 1200-7.
- Adachi T, Pimentel DR, Heibeck T, et al (2004). S-glutathiolation of Ras mediates redox-sensitive signaling by angiotensin II in vascular smooth muscle cells. *J Biol Chem*, **279**, 29857-62.
- Adjei A (2001). Blocking oncogenic ras signaling for cancer therapy. *J Natl Cancer Inst*, **93**, 1062-74.
- Ahmad N, Adhami NM, Afaq F, Feyes DK, Mukhar H (2001). Resveratrol causes WAF-1/P21-mediated G(1)-phase arrest of cell cycle and induction of apoptosis in human epidemoid carcinoma A431 cells. *Clin Cancer Res*, **7**, 1466-73.
- Ahsan N, Ali A, Ali R (2003). Oxygen free radicals and systemic autoimmunity. *Clin Exp Immunol*, **31**, 394-404.
- Ames BN, Gold LS, Willet WC (1994). Causes and prevention of cancer. *Proc Natl Acad Sci USA*, **92**, 5258-65.
- Ames BN, Shigenaga MK, Gold LS (1993). DNA lesions, inducible DNA repair, and cell division; three key factors in mutagenesis and carcinogenesis. *Environ Health Perspect*, **101** (Supp 15), 35-44.
- Ames BN, Shigenaga MK, Gold LS (1993). DNA lesions, inducible DNA repair, and cell division; three key factors in mutagenesis and carcinogenesis. *Environ Health Perspect*, **101** (Supp 15), 35-44.
- Arora A, Kalra N, Shukla Y (2006). Regulation of P21/ras protein expression by DAS in DMBA induced neoplastic changes in mouse skin. *Cancer letters*, **42**, 28-36.
- Aruoma, OI, Halliwell B, Dizdaroglu M (1989). Iron ion-dependent modification of bases in DNA by the superoxide radical-generating system hypoxanthine/xanthine oxidase. *J Biol Chem*, **264**, 13024-8.
- Bernatchez PN, Soker S, Sirois MG (1999). Vascular endothelial growth factors effect on endothelial cell proliferation, migration, and platelet-activating factor synthesis is FIK-1 dependent. *J Biol Chem*, **274**, 31047-54.
- Bharti AC, Donato N, Singh S & Aggawal BB (2003). Curcumin (diferulomethane) down-regulate the constitutive activation of nuclear factor-KB and IKK β kinase in human multiple myeloma cells, leading to suppression of proliferation and induction of apoptosis. *Blood*, **??**, 1053-62.
- Bianchini F, Vainio H (2001). Allium vegetables and organosulphur compounds: do they help prevent cancer? *Environ Health Perspect*, **109**, 893-902.
- Bode A, Dong Z (2004). Targeting signal transduction pathways by chemopreventive agents. *Mutat Res*, **555**, 33-51.
- Boff J, Min DB (2002). Chemistry and reaction of singlet oxygen in foods. *Comp Rev Food Sci Sat*, **1**, 58-72.
- Brandes RP, Koddenberg G, Gwinner W, et al (1999). Role of increased production of superoxide anions by NAD(P)H oxidase and xanthine oxidase in prolonged endotoxemia. *Hypertension*, **33**, 1243-1249.
- Brown RK, McBurney A, Lunec J, Kelly FJ (1995). Oxidative damage to DNA in patients with cystic fibrosis. *Free Radic Biol Med*, **18**, 8101-61
- Buechter DC (1988). Free radicals and oxygen toxicity. *Pharm Res*, **5**, 253-60.
- Chen C, King A (2005). Dietary cancer-chemopreventive compounds: from signaling and gene expression to pharmacological. *Trends Mol Med*, **26**, 318-28.
- Chiao C, Carothers AM, Grunberger D, Solomon G, Preston GA, Barrett JC (1995). Apoptosis and altered state induced by CAPE in transformed rat fibroblast cells. *Cancer Res*, **55**, 3576-83.
- Chung FL, Conoway CC, Rao CV and Reddy BS (2000). Chemo crypt foci in Fisher rats by sulforaphane and phenethyl isothiocyanate. *Carcinogenesis*, **21**, 2287-91.
- Chung J, Huang C, Meng X, Dong Z, Yang C (1999). Inhibition of activator protein 1 activity and cell growth by purified green tea and black tea polyphenols in H-ras-transformed cells: structure-activity relationship and mechanisms involved. *Cancer Res*, **59**, 4610-7.
- Davis-Smyth T, Chem H, Park J, Presta LG, Ferrara N (1996). The second immunoglobulin-like domain of the VEGF tyrosine Kinase receptor Fik-1 determines ligand binding and may initiate a signal transduction cascade. *EMBO J*, **15**, 4919-27.
- De Keulenaer GW, Alexander RW, Ushio-Fukai M, Ishizaka N, Griending KK (1998). Tumour necrosis factor α activates a p22phox-based NADH oxidase in vascular smooth muscle. *Biochem J*, **329**, 653-7.
- Demling RH, Lalonde C, Jin L-J, Ryan P, Fox R (1986). Endotoxemia causes increased lung tissue lipid peroxidation in unanesthetized sheep. *J Appl Physiol*, **60**, 2094-100.
- Dimasco P, Kaiser S, Sies H (1989). Lycopene as the most effective biological carotenoid singlet oxygen quencher. *Arch Biochem Biophys*, **274**, 532-38.
- Droge W (2002). Free radicals in the physiological control of cell function. *Physiol Rev*, **82**, 47-95.
- Farombi EO, Shirotriya S, Wa HK, Kim SH, Surh YJ (2008). *Food Chem Toxicol*, **46**, 1279-87.
- Ferro A, Kefer J, Bando M, Niles W, Malik AB (1998). E-selectin expression in human endothelial cells by TNF-alpha-induced oxidant generation and NF-kB activation. *Am J Physiol Lung Cell Mol Physiol*, **275**, L533-44.
- Ferro TJ, Gertzberg N, Selden L, Neumann P, Johnson A (1997). Endothelial barrier dysfunction and p42 oxidation induced by TNF-alpha are mediated by nitric oxide. *Am J Physiol Lung Cell Mol Physiol*, **272**, L979-88.
- Finkel T (1998). Oxygen radicals and signaling. *Curr Opin Cell Biol*, **10**, 248-53.
- Finkel T, Holbrook NJ (2000). Oxidants, oxidative stress and the biology of ageing. *Nature*, **408**, 39-247.
- Forman HJ, Torres M, Fukuto J (2002). Redox signaling. *Mol Cell Biochem*, **234-5**, 49-62.
- Fulda S, Debatin KM (2006). Resveratrol modulation of signal transduction in apoptosis and cell survival: a mini review. *Cancer Detect Prev*, **30**, 217-23.
- Giovannucci E, Rimm EB, Liu Y, Stampfer MJ, Willet WC (2002). A prospective study of tomato products, lycopene and prostate cancer risk. *J Natl Cancer Inst*, **94**, 391-8.
- Goode HF, Webster NR (1993). Free radicals and antioxidants in sepsis. *Crit Care Med*, **21**, 1770-6.
- Green M, Newell O, Aboyade-Cole A, Darling-Reed S, Thomas RD (2007a). DAS induces the expression of estrogen metabolizing genes in the presence and/or absence of diethylstilbestrol in the breast of female ACI rats. *Toxicol letters*, **168**, 7-12.
- Green M, Newell O, Aboyade-Cole A, Darling-Reed S, Thomas RD (2007b). DAS induces the expression of nucleotide excision repair enzymes in the breast of female ACI rats. *Toxicol letters*, **168**, 40-4.
- Griending KK, Minieri CA, Ollerenshaw JD, Alexander RW (1994). Angiotensin II stimulates NADH and NADPH oxidase activity in cultured vascular smooth muscle cells. *Circ Res*, **74**, 1141-8.
- Griending KK, Sorescu D, Ushio-Fukai M (2000). NAD(P)H

- oxidase. Role in cardiovascular biology and disease. *Circ Res*, **86**, 494-501.
- Grunberger D, Banerjee R, Eisinger K et al. (1988). Preferential cytotoxicity on tumour cells by caffeic acid ester isolated from propolis. *Experientia*, **44**, 230-2.
- Gulam W, Haseeb A (2006). Reactive oxygen species: role in the development of cancer and various chronic conditions. *J Carcinog*, **5**, 5-14.
- Gupta S, Afaq F, Mukhtar H (2002). Involvement of nuclear factor-kappa B, Bax, and Bcl-2 in induction of cell cycle arrest and apoptosis by apigenin in human prostate carcinoma cells. *Oncogene*, **21**, 3727-38.
- Hagen TM, Huang S, Curnutte J, et al (1994). Extensive oxidative DNA damage in hepatocytes of transgenic mice with chronic active hepatitis destined to develop hepatocellular carcinoma. *Proc Natl Acad Sci USA*, **91**, 12808-12.
- Hail N (2003). Mechanisms of vanilloid-induced apoptosis. *Apoptosis*, **8**, 251-62.
- Hajra K, Liu J (2004). Apoptosome dysfunction in human cancer. *Apoptosis*, **9**, 691-704.
- Halliwell B (1993). The chemistry of free radicals. *Toxicol Ind Health*, **9**, 1-21.
- Halliwell B (1997). Antioxidants and human disease: a general introduction. *Nutr Rev*, **55**, 544-9.
- Halliwell B, Gutteridge JM (1990). Role of free radicals and catalytic metal ions in human disease: an overview methods *Enzymol*, **186**, 1-85.
- Halliwell B, Gutteridge JM (1992). Biologically relevant metal ion-dependent hydroxyl radical generation. An update. *FEBS lett*, **307**, 108-12.
- Halliwell B, Gutteridge JMC (1999). Free Radicals in Biology and Medicine. 3. Oxford: Oxford University Press;
- Han SS, Keum YS, Chun KS, Surh YJ (2002). Suppression of phorbol ester-induced NF-kb activation by capsaicin in cultured human promyelocytic leukemia cells. *Arch Pharm Res*, **25**, 475-9.
- Hanai J, Dhanabal M, Karumanchi SA, et al (2002). Endostatin causes G1 arrest of endothelial cells through inhibition of cyclin D1. *J Biol Chem*, **277**, 16464-9.
- Harms K, Nozell S, Chen X (2004). The common and distinct target genes of the p53 family transcription factors. *Cell Mol Life Sci*, **61**, 822-42.
- Harris CC, Hollstein M (1993). Clinical implications of the P53 tumor suppressor gene. *N Engl J Med*, **329**, 131-27.
- Heim KE, Tagliaferro AR, Bobilya DJ (2002). Flavonoid antioxidant: Chemistry, metabolism and structure-activity relationship. *J Nutr Biochem*, **13**, 572-84.
- Howells L, Gallacher-Horley B, Houghton C (2002). Indole-3-carbinol inhibits Akt/PKB and induces apoptosis in the human breast tumor cell line MDA MB468, but not in the nontumorigenic HBL100 line. *Mol Cancer Ther*, **1**, 1161-72.
- Hwang ES, Lee JH (2006). Inhibitory effects of lycopene on the adhesion, invasion, and migration of SK-Hep 1 human hepatoma cells. *Exp Biol Med*, **231**, 322-7.
- Hwang ES, Bowen PE (2000) Cell cycle arrest and induction of apoptosis by lycopene in LNCaP human prostate cancer cell. IARC working Group on the evaluation of cancer preventive agents (1998). IARC Handbooks of Cancer Prevention: Carotenoids, **2**, 1-326.
- Jain CK, Agarwal S, Rao AV (1999). The effect of dietary lycopene on bioavailability, tissue distribution, in-vivo antioxidant properties and colonic preneoplasia in rats. *Nutr Res*, **19**, 1383-91.
- Jang M, Pezzuto JM (1998). Effects of resveratrol on 12-O-tetradecanoyl phorbol-13-acetate-induced oxidative events and gene expression of short-term ultraviolet B radiation-mediated damages by resveratrol in SKH-1 hairless mice. *Toxicol Appl Pharmacol*, **186**, 28-37.
- Janssen-Heininger YMW, Poynter ME, Baeuerle PA (2000). Recent advances towards understanding redox mechanisms in the activation of nuclear factor [kappa]b. *Free Radic Biol Med*, **28**, 1317-27.
- Jeong KM, Kyu YH, Eok CK, et al (2004). Capsaicin inhibits in vitro and in vivo angiogenesis. *Cancer Res*, **64**, 644-51.
- Johnson T, Yu Z, Ferrans V, Lowenstein R, Finkel T (1996). Reactive oxygen species are downstream mediators of p53-dependent apoptosis. *Proc Natl Acad Sci USA*, **93**, 11848-52.
- Johnstone R, Rueflir A, Lowe S (2002). Apoptosis: a link between cancer genetics and chemotherapy. *Cell*, **108**, 153-64.
- Jung MY, Kang HJ, Moon A (2001). Capsaicin-induced apoptosis in SK- Hep-1 hepatocarcinoma cells involved Bcl-2 down regulation and caspase-3 activation. *Cancer letters*, **165**, 139-45.
- Kalayarasan S, Sriram N, Sudhandiran G (2008). Diallyl sulphide attenuates bleomycin-induced pulmonary fibrosis: critical role of INOS, NF-Kappa B, TNF-alpha and IL-1 beta. *Life Sciences*, **82** (23-24), 1142-53.
- Kassahun K, Davis M, Hu P, Martin B, Baillie T (1997) Biotransformation of the naturally occurring isothiocyanate sulforaphane in the rat: identification of phase I metabolites and glutathione conjugates. *Chem Res Toxicol*, **10**, 1228-33.
- Kim JS, Kim JM, Jung HCI, Song JS (2001). Expression of cyclooxygenase-2 in human neutrophils activated by Helicobacter pylori water-soluble proteins' possible involvement of NF-Kappa B and MAP kinase signaling pathway. *Digest Dis SA*, **46**, 2277-84.
- Kim JS, Kim JM, Jung HCI, Song JS (2001). Expression of cyclooxygenase-2 in human neutrophils activated by Helicobacter pylori water-soluble proteins' possible involvement of NF-Kappa B and MAP kinase signaling pathway. *Digest Dis SA*, **46**, 2277-84.
- Kimura Y, Okuda H, Okuda T, et al (1985). Studies on the activities of tannins and related compounds from medicinal plants and drugs VII. Effects of extracts of leaves of Artemisia species, and caffeic acid and chlorogenic acid on lipid metabolic injury in rats fed peroxidized oil. *Chem Pharm Bull*, **33**, 2028-34.
- Klaunig JE, Kamendulis LM (2004). The role oxidative stress in carcinogenesis. *Ann Rev Pharmacol Toxicol*, **44**, 239-392.
- Knight JA (1995). Diseases related to oxygen-derived free radicals. *Ann Clin Lab Sci*, **25**, 111-21.
- Kolm RH, Danielson UH, Zhang Y, Talalay P and Mannervik B (1995) Isothiocyanates as substrates for human glutathione transferases: structure-activity studies. *Biochem J*, **311**, 453-9.
- Krutvoskikh V, Asamoto M, Tukesuka N et al. (1997). Differential dose-dependent effect of A- & B-carotenes in and lycopene on gap junctional intercellular communication in rat liver in vivo. *Jpn J Cancer Res*, **88**, 1121-4.
- Kundu JK, Surh YJ (2005). Molecular basis of chemoprevention by resveratrol: NF-KB and AP-1 as potential targets. *Mutat Res*, **555**, 65-80.
- Laranjinha J, Vieria O, Madeira V, Almeida L (1995). Two related phenolic antioxidants with opposite effects on Vitamin E content in low density lipoproteins oxidized by ferrylmyoglobin: consumption vs regeneration. *Arch Biochem Biophys*, **323**, 373-81.
- Lee SR, Yang KS, Kwon J, Lee C, Jeong W, Rhee SG (2002). Reversible inactivation of the tumor suppressor PTEN by H₂O₂. *J Biol Chem*, **277**, 20336-42.
- Lee YS, Nam DH, Kim JA (2001). Induction of apoptosis by capsaicin. *Cancer letters*, **120**, 235-41.

- Levine A (1997). p53, the cellular gatekeeper for growth and division. *Cell*, **88**, 323-31.
- Li ZG, Hong T, Shimada Y, et al (2002). Suppression of N-nitrosomethylbenzylamine (NMBA)-induced esophageal tumorigenesis in F344 rats by resveratrol. *Carcinogenesis*, **23**, 1531-6.
- Lien AI, Pham-Huy, Hua H, Chuong PH (2008). Free radicals, antioxidant in disease and health. *Int J Biomed Sci*, **4**, 89-96.
- Lum H, Roebuck KA (2001). Oxidant stress and endothelial cell dysfunction. *Am J Cell Physiol*, **280**, C719-41.
- Lunec J, Holloway KA, Cooke MS, Faux S, Griffiths HR, Evans MD (2002). Urinary 8-oxo-2'-deoxyguanosine: redox regulation of DNA repair *in vivo*? *Free Rad Biol Med*, **33**, 875-85.
- Macho A, Blazquez MV, Navas P, Munoz E (1998). Induction of apoptosis by vallooid compounds does not require de novo gene transcription and activator protein 1 activity. *Cell Growth Differ*, **9**, 277-435.
- Manson M, Farmer P, Gescher A, Steward W (2005). Innovative agents in cancer prevention. *Rec Res Cancer Res*, **166**, 257-75.
- Martindale JL, Holbrook NJ (2002). Cellular response to oxidative stress signaling for suicide and survival. *J Cell Physiol*, **192**, 1-15.
- Martinez J, Moreno J (2000). Effect of resveratrol, a natural polyphenolic compound, on reactive oxygen species and prostaglandin production. *Biochem Pharmacol*, **59**, 865-70.
- Matsubara T, Ziff M (1986). Increased superoxide anion release from human endothelial cells in response to cytokines. *J Immunol*, **137**, 3295-8.
- Matsushima NR, Shidoji Y, Nishiwaki et al. (1995). Suppression by carotenoids of microsystin-induced morphological changes in mouse hepatocytes. *Lipids*, **30**, 1029-34.
- Mihara M, Erster S, Zaika A, et al (2003) p53 has a direct apoptogenic role at the mitochondria. *Mol Cell*, **11**, 577-90.
- Miller NJ, Sampson J, CAndeias LP, Bramley PM, Rice-Evans CA (1996). Antioxidant activities of carotenes and xanthophylls. *FEBS Lett*, **384**, 240-6.
- Mohammad et al. (2007) Resretatrol. A Review of pre-clinical studies for human cancer. *Toxicol Appl Pharmacol*, **224**, 274-83.
- Moller P, Wallin H (1998). Adduct formation, mutagenesis and nucleotide excision repair of DNA damage produced by reactive oxygen species and lipid peroxidation product. *Mutat Res*, **410**, 271-90.
- Motterlini R, Foresti R, Bassi R, Green CJ (2000). Curcumin, an antioxidant and anti-nflamatory agent, induces heme oxygenase-1 and protects endothelial cells against oxidative stress. *Free Radic Biol Med*, **28**, 1303-12.
- Mouad ??? et al (2005). Extracellular matrix stimulate reactive oxygen species production and increase pancreatic cancer cell survival through 5-lipoxygenase and NADPH oxidase. *Am J Physiology Gastrointest Liver Physiol*, **289**, G1137-47.
- Mueller CF, Laude K, McNally JS, Harrison DG (2005). ATVB in focus: redox mechanisms in blood vessels. *Arterioscler Thromb Vasc Biol*, **25**, 274-78.
- Mukhopadhyay A, Banerjee S, Stafford LJ et al. (2002). Curcumin induced Suppression of cell proliferation correlations in the down-regulation of cyclin D1 expression and CDK4-mediated retinoblastoma protein phosphorylation. *Oncogene*, **21**, 8852-61.
- Murphy , et al (1992). Superoxide responses of endothelial cells to C5a and TNF-alpha: divergent signal transduction pathways. *Am J Physiol Lung Cell Mol Physiol*, **263**, L51-9.
- Narisawa T, Fukaura Y, Hasebe M et al. (1998). Prevention of N-methyl nitrosourea-induced colon carcinogenesis in F344 rats by lycopene. *Jpn J Cancer Res*, **89**, 1003-8.
- Natarajan K, Singh S, Burke TR Jr, Grunberger D and Aggarwal BB (1996). Caffeic acid phenethyl ester is a potent and specific inhibitor of activation of nuclear transcription factor NF-kappa B. *Proc Natl Acad Sci USA*, **93**, 9090-5.
- Natarajan K, Singh S, Burke TR Jr, Grunberger D, Aggarwal BB (1996). CAPE is a potent and specific inhibitor of activation of nuclear transcription factor NF-KB. *Proc Natl Acad Sci USA*, **93**, 9090-5.
- Okajima E, Tsutsumi M, Ozono S, et al (1998). Inhibitory effect of tomato juice on rat urinary bladder carcinogenesis alter N-butyl-N-(4 hydroxyl) nitrosamine initiation. *Jpn J Cancer Res*, **89**, 2-26.
- Olinski R, Jaruga P, Zastawny TH (1998). Oxidative DNA base modifications as factors in carcinogenesis. *Acta Biochem Pol*, **45**, 561-72.
- Orban Z, Mitsaides N, Burke TR Jr, Isokos M, Chrousos GP (2000). CAPE induces leukocyte apoptosis, modulates nuclear factor-kappa B and suppresses acute inflammation. *Neuroimmunomodation*, **7**, 99-105.
- Ozyurt H, Sogut S, Yildirim Z, et al (2004). Inhibitory effect of caffeic acid phenethyl ester on bleomycine-induced lung fibrosis in rats. *Clin Chim Acta*, **339**, 65-75.
- Palozza P (2002). Pro-oxidant and autiodant mechanism(s) of BHT and B-carotene in photocarcinogenesis. *Front Biosci*, **7**, 1044-55.
- Park HS, Jung HY, Park EY, et al (2004). Cutting edge: direct interaction of TLR4 with NAD(P)H oxidase 4 isozyme is essential for lipopolysaccharide-induced production of reactive oxygen species and activation of NF-kB. *J Immunol*, **173**, 3589-93.
- Park KK, Chun KS, Yook JI, Surh YJ (1998). Lack of tumour promoting activity of capsaicin, a principal pungent ingredient of red pepper, in mouse skin carcinogenesis. *Anticancer Res*, **18**, 4201-4205.
- Phelps DT, Ferro TJ, Higgins PJ, et al (1995). TNF-alpha induces peroxynitrite-mediated depletion of lung endothelial glutathione via protein kinase C. *Am J Physiol*, **269**, L551-9.
- Philip S, Kindu GO (2003). Osteopontin induces nuclear factor KB, medicated promatrix metalloproteinase -2 activation through 1KB & /1KK signaling pathways, and curcumin (diferulomethane) down regulate theses pathways. *J Biol Chem*, **278**, 14487-97.
- Pincemail J (1997). Free radicals and antioxidants in human disease. In favier AE, Cadet J, Kalyanaraman B, Fontecare M, Pierre-J.L (eds). "Analysis of Free Radicals in Biological Systems". Basal, Switzerland: Birkhauser Verlagin Pp. 83-99.
- Poli G, Leonarduzzi G, Biasi F, Chiarotto E (2004). Oxidative stress and cell signaling. *Curr Med Chem*, **11**, 1163-82.
- Pool-Zobel BL, Bub A, Muller H, Woolouski I, Rechkemmer G (1997). Consumption of vegetables reduces genetic damage in humans: first result of a human intervention trial with carotenoid-rich foods. *Carcinogenesis*, **18**, 1847-50.
- Rajesshwar AM (1996). Phytochemical strategies for abating environmental pollution. *Chem Ind*, **12**, 454-8.
- Rao AV, Agarwal S (1998). Bioavailability and in vivo antioxidant properties of lycopene from tomato products and their possible role in the prevention of cancer. *Nutr Cancer*, **31**, 199-203.
- Roemer K, Mahyar-Roemer M (2002). The basis for the chemopreventive action of resveratrol. *Drugs Today*, **38**, 571-80.
- Saitoh M, Nishitoh H, Fujii M, et al (1998). Mammalian thioredoxin is a direct inhibitor of apoptosis signal-regulating kinase (ASK) 1. *EMBO J*, **17**, 2596-606.

- Salvemini D, Cuzzocrea S (2002). Oxidative stress in septic shock and disseminate intravascular coagulation. *Free Radical Biol Med*, **33**, 1173-85.
- Schafer FQ, Buettner GR (2001). Redox environment of the cell as viewed through the redox state of the glutathione disulfide/glutathione couple. *Free Radic Biol Med*, **30**, 1191-212.
- Schwartz JL (1992). The dual roles of nutrients as antioxidants. *Nutrition*, **17**, 815-7.
- Sesso HD, Buring JE, Norkus EP, Gaziano JM (2004). Lycopene other carotenoids and retinol and the risk of cardiovascular disease in women. *Am J Clin Nutr*, **79**, 47-53.
- Sharoni Y, Giron E, Rise M, Levy J (1997). Effects of lycopene-enriched tomato oleoresin on 7, 12-dimethylbenz[a]anthracene-induced rat mammary tumours. *Cancer Detect Prev*, **21**, 118-123.
- Shimoda R, Nagashima M, Sakamoto M, et al (1994). Increased formation of oxidative DNA damage 8-hydroxydeoxyguanosine, in human livers with chronic hepatitis. *Cancer Res*, **54**, 3171-2.
- Slee E, O'Connor D, Lu X (2004). To die or not to die: how does p53 decide? *Oncogene*, **23**, 2809-818.
- Song YS, Park EH, Hur GM, et al (2002). Caffeic acid phenethyl ester inhibits nitric oxide synthase gene expression and enzyme activity. *Cancer Lett*, **175**, 53-61.
- Spiteller G (2001). Lipid oxidation in aging and age-dependent disease. *Exp Gerontol*, **36**, 1425-57.
- Sriram N, Kalayarasan S, Ashokkumar P, Sudhandiran G (2008). DAS induces apoptosis in colon 320 DM human colon cancer cells: involvement of caspase-3, NF-Kappa B, and ERK-2. *Mol Cell Biochem*, **311**, 157-65.
- Stahl W, Junghans B, Boer ES, et al (1998) Carotenoid mixtures protect multilamellar liposomes against oxidative damage: synergistic effects of lycopene and lutein. *FEBS letters*, **427**, 305-8.
- Subbaramaiah K, Chung WJ, Michaluart P, et al (1998). Resveratrol inhibits cyclooxygenase-transcription and activity in phorbol ester-treated human mammary epithelial cells. *J Bio Chem*, **273**, 21875-882.
- Sun S, Hail N, Lotan R (2004). Apoptosis as a novel target for cancer chemoprevention. *J Natl Cancer Inst*, **96**, 662-72.
- Sundaresan M, Yu Z-X, Ferrans VJ, Irani K, Fingel T (1995). Requirement for generation of H₂O₂ for platelet-derived growth factor signal transduction. *Science*, **270**, 296-9.
- Surh YJ (2002). More than spice: Capsaicin in hot chili peppers makes tumour cells commit suicide. *J Natl Cancer Inst*, **94**, 1263-5.
- Surh YJ, Lee SS (1995). Capsaicin, a double-edged sword: toxicity, metabolism, and chemopreventive potential. *Life Sci*, **56**, 1845-55.
- Szatrowski TP, Nathan CF (1991). Production of large amount of hydrogen peroxide by human tumour cells. *Cancer Res*, **51**, 794-8.
- Thannickal VJ, Fanburg BL (2000). Reactive oxygen species in cell signalling. *Am J Physiol Lung Cell Mol Physiol*, **279**, L1005-28.
- Thejass P, Kuttan G (2007). Antiangiogenic activity of diallyl sulfide (DAS). *Int Immunopharmacology*, **7**, 295-305.
- Thomas RD, Green MR, Wilson C, Sadrud-Din S (2004). DAS inhibits the oxidation and reduction reactions of stilbene estrogens catalyzed by microsomes, mitochondria and nuclei isolated from breast tissue of female ACF rats. *Carcinogenesis*, **25**, 787-91.
- Turrens JF (2003). Mitochondrial formation of reactive oxygen species. *J Physiol*, **552**, 335-44.
- Ullrich V, Bachschmid M (2000). Superoxide as a messenger of endothelial function. *Biochem Biophys Res Commun*, **278**, 1-8.
- Ushio-Fukai M, Alexander RW, Akers M, et al (1999). Reactive oxygen species mediate the activation of Akt/protein kinase B by angiotensin II in vascular smooth muscle cells. *J Biol Chem*, **274**, 22699-704.
- Wada T, Penninger JM (2004). Mitogen-activated protein kinases in a poptosis regulation. *Oncogene*, **23**, 2838-49.
- Waris G, Alam K (1998). Attenuated antigenicity of ribonucleoproteins modified by reactive oxygen species. *Biochem Mol Biol Int*, **45**, 3345.
- Waris R, Turkson J, Hassanein T, Siddiqui A (2005). Hepatitis C virus constitutively activates STAT-3 via oxidative stress: Role of STAT-3 in HCV replication. *J Virol*, **79**, 1569-80.
- White CR, Darley-Usmar V, Berrington WR, et al. (1996). Circulating plasma xanthine oxidase contributes to vascular dysfunction in hypercholesterolemic rabbits. *Proc Natl Acad Sci USA*, **93**, 8745-9.
- Witztum JL (1994). The oxidation hypothesis of atherosclerosis. *Lancet*, **344**, 793-5.
- Woodall AA, Britton G, Jackson MH (1997). Carotenoids and protection of phospholipids in solution or in liposomes against oxidation by peroxy radicals: relationship between carotenoid structure and protective ability. *Biochem Biophys Acta*, **1336**, 575-86.
- Woods D, Vousden K (2001). Regulation of p53 function. *Exp Cell Res*, **264**, 56-66.
- Wu X, Kassie F, Mersch-Sundermann V (2005). Induction of apoptosis in tumor cells by naturally occurring sulfur-containing compounds. *Mutat Res*, **589**, 81-102.
- Xia Y, Dawson VL, Dawson TM, Snyder SH, Zweier JL (1996). Nitric oxide synthase generates superoxide and nitric oxide in arginine-depleted cells leading to peroxynitrite-mediated cellular injury. *Proc Natl Acad Sci USA*, **93**, 6770-4.
- Xia Y, Zweier JL (1997). Superoxide and peroxynitrite generation from inducible nitric oxide synthase in macrophages. *Proc Natl Acad Sci USA*, **94**, 6954-8.
- Ye L, Zhang Y (2002). Total intracellular accumulation levels of dietary isothiocyanates determine their activity in elevation of cellular glutathione and induction of phase 2 detoxication enzymes. *Carcinogenesis*, **22**, 1987-92.
- Zachary I, Glick G (2001). Signaling transduction mechanism mediating biological actions of the vascular endothelial growth factor family. *Cardiovasc Res*, **49**, 568-81.
- Zafari et al. (1998). Role of NADH/NADPH oxidase-derived H₂O₂ in angiotensin II-induced vascular hypertrophy. *Hypertension*, **32**, 488-95.
- Zhang Y (2000). Role of glutathione in the accumulation of anticarcinogenic isothiocyanates and their glutathiones by murine hepatoma cells. *Carcinogenesis*, **21**, 1175-82.
- Zhang Y (2001). Molecular mechanisms of rapid cellular accumulation of anticarcinogenic isothiocyanates. *Carcinogenesis*, **22**, 425-31.
- Zhang Y (2004). Cancer-preventive isothiocyanates: measurement of human exposure and mechanism of action. *Mutat Res*, **555**, 173-90.
- Zhang Y, Talalay P (1998). Mechanism of differential potencies of isothiocyanates as inducers of anti-carcinogenic Phase 2 enzymes. *Cancer Res*, **58**, 4632-9.
- Zhang Y, Talalay P, Cho CG, Posner GH (1992). A major inducer of anticarcinogenic protective enzymes from broccoli: isolation and elucidation of structure. *Proc Natl Acad Sci USA*, **89**, 2399-403.